

DIETHYLAMINE



Method no.: 41

Matrix: Air

Target concentration: 10 ppm (29.9 mg/m³) (ACGIH TLV)

Procedure: Samples are collected by drawing known volumes of air through standard size sampling tubes containing XAD-7 resin coated with 10% NBD chloride by weight. The samples are desorbed with 5% (w/v) NBD chloride in tetrahydrofuran (with a small amount of sodium bicarbonate present), heated in a hot water bath, and analyzed by high-performance liquid chromatography using a fluorescence or visible detector.

Recommended air volume and sampling rate: 10 L at 0.2 L/min

Reliable quantitation limit: 53 ppb (160 µg/m³)

Standard error of estimate at the target concentration: (Section 4.4.) 7.1%

Status of method: Evaluated method. This method has been subjected to the established evaluation procedures of the Organic Methods Evaluation Branch.

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1. General discussion

1.1. Background

1.1.1. History

The recommended sampling and analytical procedure for diethylamine (DEA) used by OSHA has been collection in midget impingers containing sulfuric acid and analysis by gas chromatography (Ref. 5.1.). Impingers are cumbersome to use in the field and the analysis of free low molecular weight amines is difficult by gas chromatography (Ref. 5.2.). Thus, it was desirable to find better alternative sampling and analytical procedures for DEA.

Collection of DEA on a solid sorbent was considered. In NIOSH methods 221 and S139 (Refs. 5.3. and 5.4.) silica gel is recommended for collection of DEA in air. It was later found by NIOSH that low molecular weight amines are not stable after being collected on silica gel (Ref. 5.5.).

In this evaluation, it was found that DEA could be collected on a sampling tube containing XAD-7 resin coated with 10% NBD chloride (7-chloro-4-nitrobenzo-2-oxa-1,3-diazole) by weight. A stable derivative is formed on the coated resin.

The derivative is extracted with 5% (w/v) NBD chloride in tetrahydrofuran (THF) and analyzed by high-performance liquid chromatography. Similar procedures have been successfully evaluated for dimethylamine (OSHA Method 34) (Ref. 5.6.), ethylamine (OSHA Method 36) (Ref. 5.7.) and methylamine (OSHA Method 40) (Ref. 5.8.) and may be applicable for other volatile aliphatic amines.

The OSHA PEL is currently 25 ppm, but ACGIH has recently revised their TLV to 10 ppm. This was the basis for selection of 10 ppm as the target concentration used in this evaluation.

1.1.2. Toxic effects (This section is quoted directly from "Occupational Health Guidelines for Chemical Hazards" (Ref. 5.9.) and is for information only and should not be taken as the basis of OSHA policy.)

"Diethylamine vapor is a severe irritant to mucous membranes, eyes, and skin. Rabbits repeatedly exposed to 50 ppm for 7 hours daily showed corneal damage and pulmonary irritation. In one reported human case, liquid splashed into the eye caused immediate intense pain. In spite of emergency irrigation and treatment, the cornea became swollen and cloudy. Some permanent visual impairment resulted. Contact with the liquid causes vesiculation (blisters) and necrosis (death of tissue) of the skin."

1.1.3. Potential workplace exposure

Following are some common operations in which exposure to DEA may occur as reported in "Occupational Health Guidelines for Chemical Hazards." (Ref. 5.9.)

Diethylamine is used:

in preparation of textile finishing agents, surfactants, rubber processing chemicals, agricultural chemicals, and pharmaceuticals.

as a corrosion inhibitor in iron, steel, and metal industries.

as a polymerization inhibitor and catalyst in the polymer industry and as an intermediate in the dye industry.

as a depilatory of animal skins and in electroplating solutions.

1.1.4. Physical properties (Ref. 5.9. unless stated otherwise)

molecular weight:	73.14
boiling point:	55.5°C (760 mm Hg)
color:	colorless liquid
specific gravity:	0.71 (water = 1)

formula:	(C ₂ H ₅) ₂ NH
vapor pressure at 20°C:	195 mm Hg
flash point (closed cup):	less than -18°C
odor:	fish, ammonia-like
flammable limits in air, % by volume:	lower: 1.8; upper: 10.1
autoignition temperature:	312°C
synonyms:	diethamine; N-ethyle-ethanamine (Ref. 5.10.)

1.2. Limit defining parameters (The DEA air concentrations listed throughout this method are based on an air volume of 10 L and a solvent desorption volume of 2 mL.)

1.2.1. Detection limit of the analytical procedure

The detection limit of the analytical procedure is 8.7 ng per injection. This is the amount of DEA which will give a peak whose height is approximately five times baseline noise. (Section 4.1.)

1.2.2. Detection limit of the overall procedure

The detection limit of the overall procedure is 1.6 µg per sample (53 ppb or 160 µg/m³). This is the amount of DEA spiked on the sampling device which allows recovery of an amount of DEA equivalent to the detection limit of the analytical procedure. (Section 4.2.)

1.2.3. Reliable quantitation limit

The reliable quantitation limit is 1.6 µg per sample (53 ppb or 160 µg/m³). This is the smallest amount of DEA which can be quantitated within the requirements of a recovery of at least 75% and a precision (1.96 SD) of ±25% or better. (Section 4.2.)

The reliable quantitation limit and detection limits reported in the method are based upon optimization of the instrument for the smallest possible amount of analyte. When the target concentration of an analyte is exceptionally higher than these limits, they may not be attainable at the routine operating parameters.

1.2.4. Sensitivity

The sensitivity of the analytical procedure over a concentration range representing 0.5 to 2 times the target concentration is 290 area units per µg DEA/mL. This is determined by the slope of the calibration curve (Section 4.3.). The sensitivity will vary with the particular instrument used in the analysis.

1.2.5. Recovery

The recovery of DEA from samples used in a 15-day storage test remained above 91% when the samples were stored at refrigerated or ambient temperature. (Section 4.4.) The recovery of DEA from the collection medium during storage must be 75% or greater.

1.2.6. Precision (analytical method only)

The pooled coefficient of variation obtained from replicate determinations of analytical standards at 0.5, 1, and 2 times the target concentration is 0.006. (Section 4.3.)

1.2.7. Precision (overall procedure)

The precision at the 95% confidence level for a 15-day storage test is ±13.8%. This includes an additional ±5% for sampling error. (Section 4.4.) The overall procedure must provide results at the target concentration that are ±25% or better at the 95% confidence level.

1.2.8. Reproducibility

Six samples collected from a controlled test atmosphere, and a draft copy of this procedure were given to a chemist unassociated with this evaluation. The samples were analyzed

after five days of storage at ambient conditions. The average recovery (corrected for desorption efficiency) was 99.9% with a standard deviation of $\pm 3.5\%$. (Section 4.5.)

1.3. Advantages

- 1.3.1. The solid sorbent tube provides a convenient method for sampling.
- 1.3.2. DEA is analyzed as a derivative which is specific, stable, and easier to quantitate than the free amine.
- 1.3.3. The analysis is rapid, sensitive, and precise.
- 1.3.4. This method is applicable to methylamine, dimethylamine, and ethylamine, and possibly other volatile aliphatic amines.

1.4. Disadvantages

- 1.4.1. The method has not been field tested.
- 1.4.2. Sampling tubes (XAD-7 coated with 10% NBD chloride) are commercially available from SKC as catalog no. 229-93 as of 2010.

2. Sampling Procedure

2.1. Apparatus

- 2.1.1. Samples are collected by use of a personal sampling pump that can be calibrated to within $\pm 5\%$ of the recommended flow rate with the sampling tube in line.
- 2.1.2. Samples are collected on solid sorbent tubes containing XAD-7 coated with 10% NBD chloride by weight. The tube contains two sections of coated XAD-7 resin separated by a glass wool plug. The front section contains 80 mg of coated sorbent and the back section 40 mg. The sections are held in place with glass wool plugs in a glass tube 4-mm i.d. \times 70-mm length.

The coated XAD-7 is prepared by rinsing the 20/50 mesh resin several times with methyl alcohol to remove fines. The resin is extracted for 24 h with methyl alcohol and dried by vacuum. The dried resin is then coated with 10% NBD chloride by weight using methylene chloride as a solvent. The solvent is removed by rotary evaporation.

2.2. Reagents

None required

2.3. Technique

- 2.3.1. Connect the sampling tube to the sampling pump with flexible tubing. Air being sampled should not pass through any hose or tubing before entering the sampling tube.
- 2.3.2. Place the sampling tube vertically in the employee's breathing zone.
- 2.3.3. After sampling, seal the tubes immediately with plastic caps.
- 2.3.4. Submit at least one blank for each sample set. The blank should be handled in the same manner as samples, except no air is drawn through it.
- 2.3.5. Record sample volume (in liters of air) for each sample, along with any potential interferences.
- 2.3.6. Ship any bulk sample(s) in a separate container(s) from the air samples.

2.4. Breakthrough

The average 5% breakthrough volume from a test atmosphere (air at about 80% relative humidity) containing 20.5 ppm (61.3 mg/m^3) DEA was determined to be 27.1 L. This corresponds to a loading

of 1.66 mg of DEA on the sampling tube. The sampling rate was approximately 0.2 L/min and the test atmosphere was at 21 °C and 642.5 mm Hg. (Section 4.6.)

2.5. Desorption efficiency

2.5.1. The desorption efficiency of DEA from spiked samples was 96.5% over the range of 5 to 20 ppm. (Section 4.7.)

2.5.2. The desorption efficiency must be determined for each lot of coated XAD-7.

2.6. Recommended air volume and sampling rate

2.6.1. The recommended air volume is 10 L.

2.6.2. The recommended sampling rate is 0.2 L/min.

2.7. Interferences (sampling)

2.7.1. An interference study was performed in which 10-L air samples of a test atmosphere containing approximately 10 ppm each of methylamine, dimethylamine, ethylamine, and diethylamine were collected. The test atmosphere was at approximately 80% relative humidity. There was no difference in the amount of DEA derivative recovered whether the other amines were present or not.

2.7.2. It is not known if any compound(s) will interfere with the collection of DEA on coated XAD-7 tubes.

2.7.3. Suspected interferences should be reported to the laboratory with submitted samples.

2.8. Safety precautions (sampling)

2.8.1. Attach the sampling equipment to the employee so that it will not interfere with work performance or safety.

2.8.2. Follow all safety procedures that apply to the work area being sampled.

3. Analytical procedure

3.1. Apparatus

3.1.1. High-performance liquid chromatograph (HPLC) equipped with a fluorescence and/or visible detector. For this evaluation a Schoeffel FS 970 LC Fluorometer and a Waters 440 detector were used in series with two Waters M-6000A pumps.

3.1.2. An HPLC column capable of separating the DEA derivative from NBD chloride and any interferences. A Radial CN column was used in this study in the normal phase mode since the NBD chloride derivatives fluoresce stronger in non-aqueous solvent systems. Reduced sensitivities will be obtained with the use of an aqueous reverse phase column system.

3.1.3. An electronic integrator or some other suitable method of measuring peak areas or heights. A Hewlett-Packard 3354 B/C Data System was used in this evaluation.

3.1.4. A mechanical shaker.

3.1.5. A hot water bath.

3.1.6. Volumetric flasks for preparing standards and making dilutions.

3.1.7. Pipets and syringes for preparing standards, making dilutions, and dispensing reagents.

3.1.8. Small vials with Teflon-lined caps capable of holding 3 mL.

3.2. Reagents

3.2.1. HPLC grade isopropanol and isooctane.

- 3.2.2. Reagent grade tetrahydrofuran (THF).
 - 3.2.3. Reagent grade sodium bicarbonate.
 - 3.2.4. Diethylamine. Lot 2029EH from Aldrich was used in this evaluation.
 - 3.2.5. Reagent grade NBD chloride. (7-chloro-4-nitrobenzo-2-oxa1,3-diazole). Aldrich Lot 1003 DH was used in this study.
 - 3.2.6. Desorption reagent: 5 g of NBD chloride per 100 mL of THF.
- 3.3. Standard preparation
- 3.3.1. Prepare a stock standard of DEA by diluting a known volume of DEA with THF.
 - 3.3.2. Prepare working standards by injecting microliter amounts of the stock standard into 2.0 mL of desorption reagent in a small vial. Example: If a 5- μ L aliquot of a stock standard at a concentration of 60 μ g/ μ L is injected into a vial containing 2.0 mL of desorption reagent, the working standard is equal to 300 μ g/sample (5×60) if the samples are desorbed with 2.0 mL. For a 10-L air sample this is equivalent to 30.0 mg/m³ (10.0 ppm), uncorrected for desorption efficiency.
 - 3.3.3. Add approximately 25 mg of solid sodium bicarbonate to each vial and seal with Teflon-lined caps. (This is easily done by using the large end of a standard size disposable dropping pipette as a spatula). The standards are shaken for 0.5 h and then heated for 2.5 h at 60°C in a water bath. Allow standards to cool to room temperature before analyzing.
- 3.4. Sample preparation
- 3.4.1. Transfer each section of the sample to separate vials. The glass wool plugs must be added to the vials if they contain entrapped XAD-7 beads. The glass tube is discarded.
 - 3.4.2. Add 2.0 mL of the desorption reagent to each vial.
 - 3.4.3. Add approximately 25 mg of solid sodium bicarbonate to each vial.
 - 3.4.4. Seal the vials with Teflon-lined caps and shake in a horizontal position for 0.5 h. The vials should be positioned parallel to the shaker's movements.
 - 3.4.5. Heat the vials for 2.5 h in a water bath at 60°C. Allow samples to cool to room temperature before analyzing.
- 3.5. Analysis
- 3.5.1. HPLC conditions

fluorescence detector:	465 nm excitation 525 nm emission
injection size:	15 μ L
column:	Waters Radial CN
solvent:	isooctane: isopropanol, 80:20 at 3 mL/min
retention time:	3.75 min
alternate detector:	visible at 465 nm
chromatograms:	Section 4.8.
 - 3.5.2. Peak areas (or heights) are measured by an integrator or other suitable means.
 - 3.5.3. A calibration curve is constructed by plotting peak areas (or heights) of standard injections versus μ g DEA per sample. Sample concentrations must be bracketed by standards.
- 3.6. Interferences (analytical)
- 3.6.1. Any compound that has the same general retention time as the DEA derivative and responds with the detector used is an interference. Possible interferences should be reported to the laboratory with submitted samples by the industrial hygienist. The

derivatives of ethylamine, methylamine, and dimethylamine can be separated from the DEA derivative.

3.6.2. HPLC parameters (i.e. solvent composition, column, detector, etc.) may be changed to possibly circumvent interferences.

3.6.3. Retention time on a single column is not considered proof of chemical identity. Samples over the PEL should be confirmed by GC/MS or other suitable means.

3.7. Calculations

The DEA concentration is obtained from
$$\frac{\text{mg}}{\text{m}^3} = \frac{(\text{blank corrected micrograms of DEA per sample})}{(\text{liters of air sampled})(\text{desorption efficiency})}$$

the calibration curve in terms of micrograms per sample. The air concentration for samples is calculated using the following formulae. If any DEA is found on the backup section, it is added to the amount found on the front section. This total amount is then corrected by subtracting the total amount found in the blank.

$$\text{ppm} = \frac{(\frac{\text{mg}}{\text{m}^3})(24.46)}{(73.14)} = \frac{\text{mg}}{\text{m}^3} (0.3344)$$

where 24.46 = molar volume (liters) at 25°C, 760 mm Hg
73.14 = MW of DEA

3.8. Safety precautions (analytical)

3.8.1. Avoid skin contact and inhalation of all chemicals used, especially DEA and NBD chloride.

3.8.2. Restrict the use of all chemicals to a fume hood if possible.

3.8.3. Wear safety glasses and a lab coat at all times while in the laboratory area.

4. Backup Data

4.1. Detection limit of the analytical procedure

The detection limit of 8.7 ng/injection was determined by making a 15- μL injection of a 0.58 $\mu\text{g}/\text{mL}$ standard. This standard is based on the weight of DEA added to make the standard. Figure 4.1. is a chromatogram obtained from a Schoeffel FS 970 Fluorescence detector set at 0.1 μA range, 4.59 sensitivity and 0.5 s time constant. A Radial CN column was used. The recorder was set at 0.2 cm/min and 10 mV full scale. The solvent system used for this determination was 90:10, isooctane:isopropanol. This was used to obtain separation between the DEA derivative and a small interference. This interference is insignificant for samples near the PEL.

4.2. Detection limit of the overall procedure and reliable quantitation limit

Six samples were prepared by spiking (liquid injection) the coated XAD-7 with 1.6 μg of DEA. The samples were analyzed the next day. The amount recovered was essentially equivalent to the analytical detection limit of 1.2 μg per sample. The results of this study are given in Table 4.2.

Table 4.2.
Detection Limit of the Overall Procedure
and Reliable Quantitation Limit Data

% recovery	statistics
72.1	
74.4	$\bar{X} = 76.8$
79.1	SD = 5.5
69.8	
81.4	
83.7	

4.3. Sensitivity and precision

The sensitivity and precision of the analytical procedure were determined from multiple injections of analytical standards. This data is given below and also shown graphically in Figure 4.3.

Table 4.3.
Sensitivity and Precision Data

x target conc. µg/mL	0.5x	1x	2x
area	25032.7	44415.1	80845.2
counts	24987.7	44830.9	81054.3
	24737.9	44973.5	80986.8
	24741.6	44783.3	80616.3
	24853.7	44593.2	80534.1
	24430.8	44894.8	79934.4
\bar{X}	24797.4	44748.5	80661.9
SD	217.2	207.5	410.1
CV	0.0088	0.0046	0.0051
$\overline{CV} = 0.006$			

4.4. Recovery and storage

Storage samples were generated from a test atmosphere (air) containing 10.7 ppm DEA at approximately 80% relative humidity, 23°C, and 640 mm Hg. Each sample was generated by sampling the test atmosphere at approximately 0.2 L/min for 50 min, resulting in a sample volume of about 10 L. An amount of coated XAD-7 equivalent to the front section of a standard adsorbent tube (about 80 mg) was used for each sample. After sampling, the adsorbent was transferred to separate vials, capped, and stored. Six samples were analyzed immediately after generation, 15 were stored in a closed drawer at ambient temperature, and 15 were stored under refrigeration at 0°C.

Table 4.4.
Storage Tests

storage time (days)	% recovery					
	(refrigerated)			(ambient)		
0	96.2	92.2	93.6	96.2	92.2	93.6
0	93.0	92.0	93.0	93.0	92.0	93.0
2	88.1	87.7	87.6	85.9	85.5	87.2
6	100.1	96.2	101.7	97.1	101.8	93.3
8	90.0	90.8	87.2	92.9	92.6	91.0
13	97.7	98.5	97.4	98.5	96.6	97.1
15	87.8	85.4	86.0	86.8	84.5	85.5

The results of these data are shown graphically in Figures 4.4.1. and 4.4.2.

4.5. Reproducibility

Six DEA samples were generated on the vapor generator by sampling from an air stream containing 10.7 ppm DEA for 50 min at approximately 0.2 L/min. The sample stream was at 25.9°C, 641.5 mm Hg, and approximately 80% relative humidity. The samples were stored for five days at room temperature before being analyzed.

Table 4.5.
Reproducibility

sample no.	mg found	mg expected	% found
1	0.3346	0.3194	104.8
2	0.3557	0.3208	100.9
3	0.3212	0.3208	100.1
4	0.3221	0.3230	99.7
5	0.3150	0.3151	100.0
6	0.3287	0.3500	93.9
7 (blank)	N.D.	0.00	---
$\bar{X} = 99.9$			
SD = 3.5			

4.6. Breakthrough

The 5% breakthrough volume was determined from a test atmosphere containing 20.5 ppm (61.3 mg/m³) DEA. The sampling tube contained only the front section (approximately 80 mg) of adsorbent. A backup tube was connected downstream from the sampling tube. This backup tube was changed periodically and analyzed to determine the amount of DEA breaking through the sampling tube. The average 5% breakthrough volume for three separate determinations was 27.1 L. This corresponds to an average loading of 1.66 mg of DEA on the sampling tube when 5% breakthrough occurred. The test atmosphere was at 21°C, 642.5 mm Hg, and approximately 80% relative humidity. A breakthrough curve for one of the tests is shown in Figure 4.6.

4.7. Desorption efficiency

The desorption efficiency was determined by injecting known amounts of a DEA standard onto six samplers of coated XAD-7 and analyzing the samples the next day.

Table 4.7.
Desorption Efficiency

$\mu\text{g/sample}$	145	290	580
ppm^1	4.85	9.69	19.4
desorption efficiency,	94.5	100.0	90.6
%	96.6	100.0	98.5
	95.0	99.7	97.1
	96.4	101.0	83.8
	97.6	99.6	96.1
	97.6	100.2	93.1
\bar{X}	96.3	100.1	93.2

$\bar{X} = 96.5$

¹air conc. for a 10 L air sample

4.8. Chromatograms

Chromatograms of a DEA standard are shown in Figure 4.8. The chromatograms are from a 15- μ L injection of a 0.291 mg of DEA per sample standard. The fluorescence and visible detectors were connected in series to give essentially simultaneous chromatograms. The responses shown are standardized to keep the peaks about 90% full scale. Thus, this figure does not indicate the relative response of each detector.

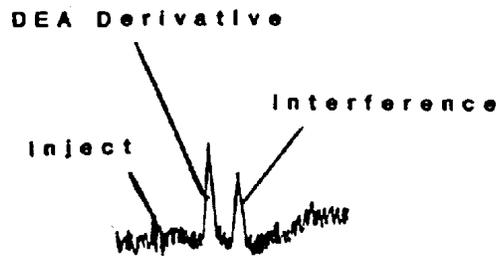


Figure 4.1. Chromatogram of the analytical detection limit.

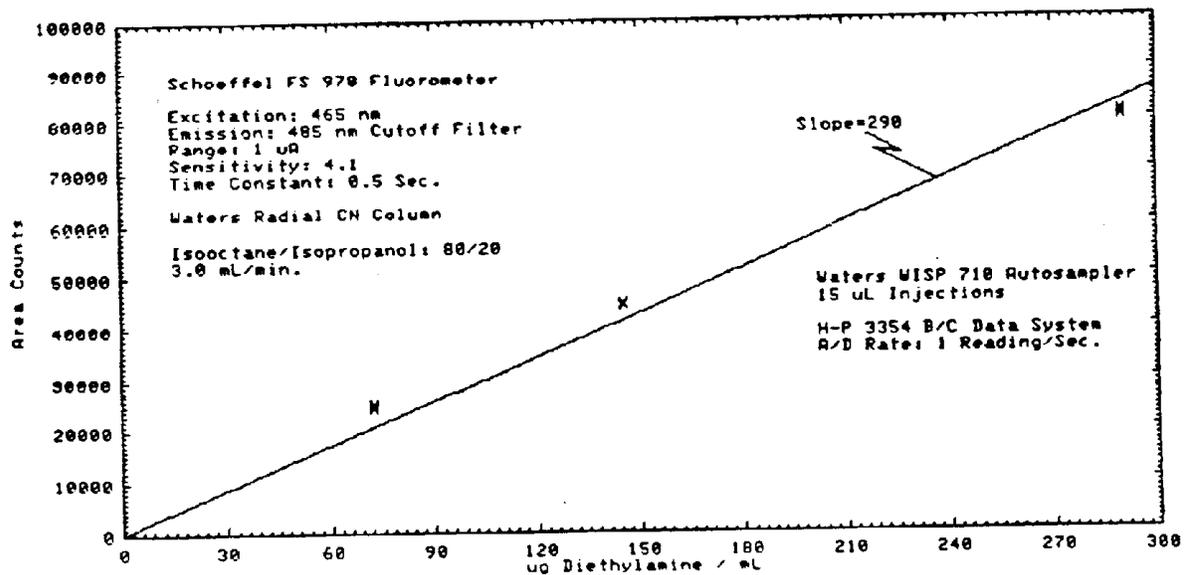


Figure 4.3. Calibration curve.

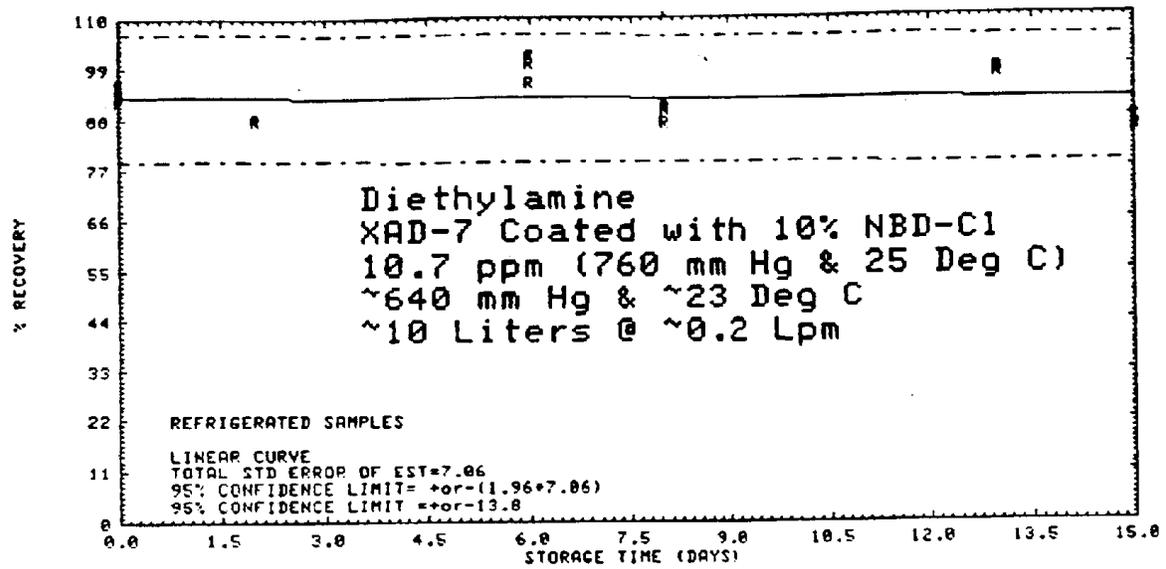


Figure 4.4.1. Refrigerated storage samples.

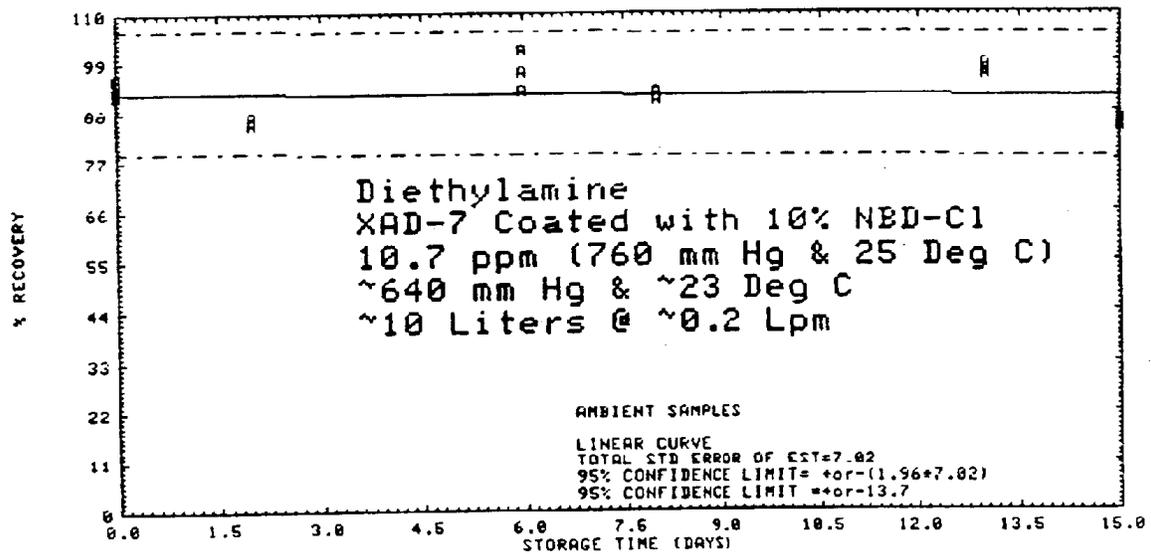


Figure 4.4.2. Ambient storage samples.

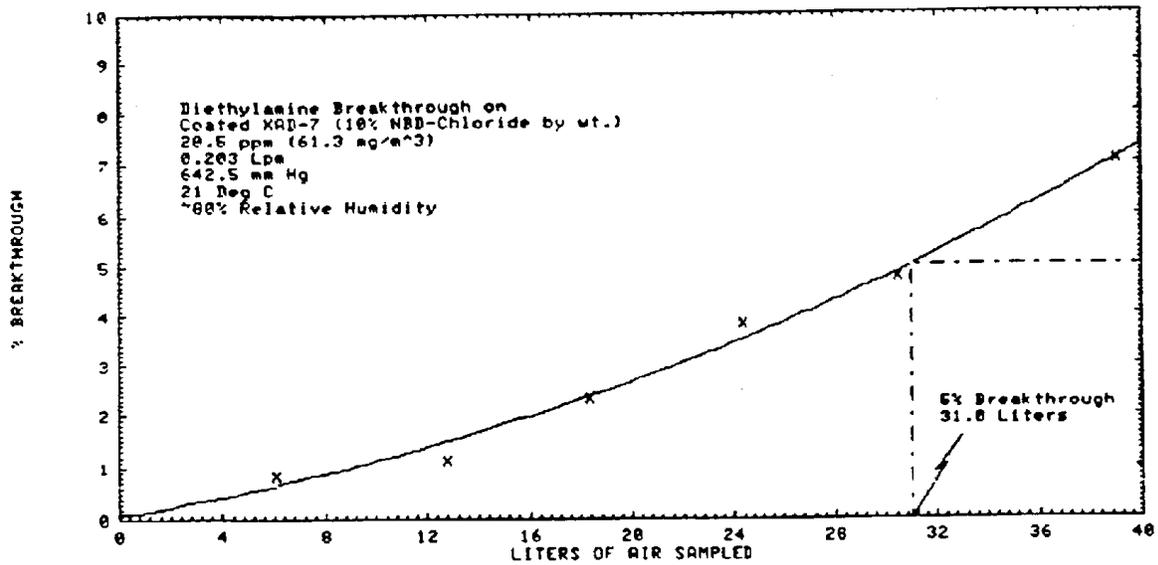


Figure 4.6. Breakthrough curve.

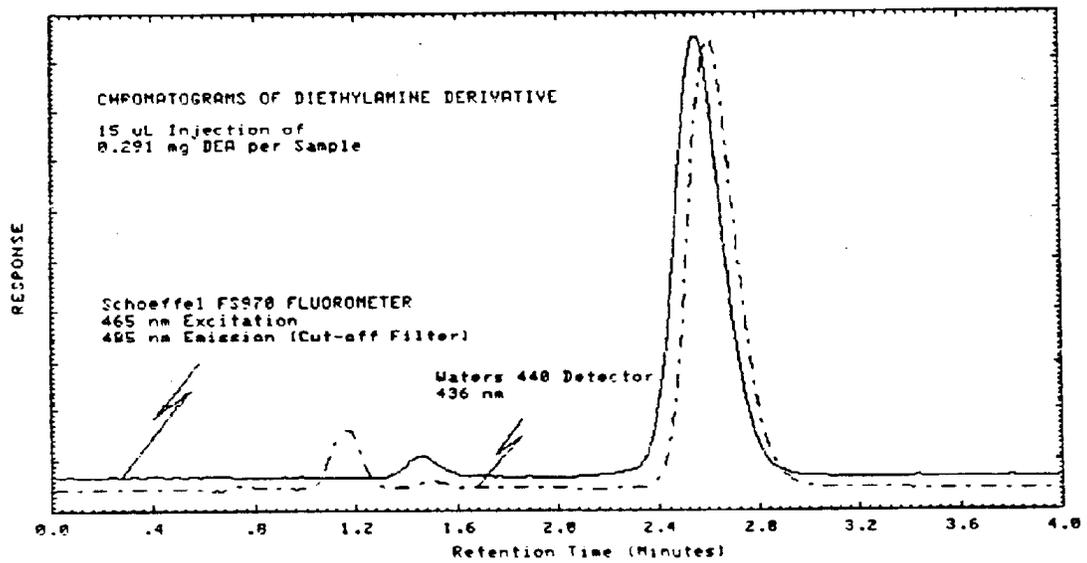


Figure 4.8. Chromatograms of a standard.

5. References

- 5.1. "Industrial Hygiene Field Operation Manual", OSHA Instruction CPL 2-2.20, Office of Field Coordination, 1979.
- 5.2. Dalene, M.; Mathiasson, L.; Jonsson, J.A. J. Chromatogr. (1981), 207, 37-46.
- 5.3. "NIOSH Manual of Analytical Methods", 2nd ed.; Department of Health, Education and Welfare, National Institute for Occupational Safety and Health: Cincinnati, OH 1977; Vol. 1, Method No. P&CAM 221; DHEW (NIOSH) Publ. (U.S.), No. 77-157-A.
- 5.4. "NIOSH Manual of Analytical Methods", 2nd ed.; Department of Health, Education and Welfare, National Institute for Occupational Safety and Health: Cincinnati, OH, 1977; Vol. 3, Method No. S139; DHEW (NIOSH) Publ. (U.S.), No. 77-157-C.
- 5.5. Teass, A. National Institute for Occupational Safety and Health, personal communication, 1981.
- 5.6. Elskamp, C.J., Dimethylamine (Method 34, Organic Methods Evaluation Branch, OSHA Analytical Laboratory, Salt Lake City, Utah). Unpublished (2-82).
- 5.7. Elskamp, C.J., Ethylamine (Method 36, Organic Methods Evaluation Branch, OSHA Analytical Laboratory, Salt Lake City, Utah). Unpublished (5-82).
- 5.8. Elskamp, C.J., Methylamine (Method 40, Organic Methods Evaluation Branch, OSHA Analytical Laboratory, Salt Lake City, Utah). Unpublished (10-82).
- 5.9. "Occupational Health Guidelines for Chemical Hazards" NIOSH/OSHA, Jan. 1981, DHHS (NIOSH) Publication No. 81-123.
- 5.10. Procter, N.H.; Hughes, J.P. "Chemical Hazards of the Workplace", J.B. Lippincott Company; Philadelphia, 1978: p 217.