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Method Number: ID-200

Matrix: Air

OSHA Permissible Exposure Limits:
Final Rule Limits: 2 ppm Time Weighted Average (TWA)

5 ppm Short-Term Exposure Limit (STEL)

Transitional Limit: 5 ppm TWA

Collection Device: An air sample is collected using a calibrated sampling pump and a glass tube containing impregnated activated beaded carbon (IABC). A prefilter/cassette assembly can be used to collect particulate, if necessary.

Recommended Sampling Rate:
TWA & STEL: 0.1 liter per minute (L/min)

Recommended Air Volume:
TWA: 12 L (0.1 L/min for 120 min)
STEL: 1.5 L (0.1 L/min for 15 min)

Analytical Procedure: The sampling medium is desorbed in 15 mM sodium hydroxide which contains 0.3 N (~1%) hydrogen peroxide. An aliquot of this solution is analyzed as sulfate by ion chromatography.

Detection Limit:
Qualitative: 0.004 ppm (12-L air sample)
0.032 ppm (1.5-L air sample)
Quantitative: 0.013 ppm (12-L air sample)
0.104 ppm (1.5-L air sample)

| Precision and Accuracy: | TWA | STEL |
|---------------------------|------------------|--------------------------|
| Validation Range: | 1.36 to 4.16 ppm | 5.79 ppm |
| CV _T (pooled): | 0.048 | 0.028 (CV ₂) |
| Bias: | -0.033 | +0.006 |
| Overall Error: | ±12.9% | ±6.2% |

Method Classification: Validated Method

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Date: April, 1992

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Commercial manufacturers and products mentioned in this method are
for descriptive use only and do not constitute endorsements by

USDOL-OSHA.

Similar products from other sources can be substituted.

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1. Introduction

This method describes the sample collection and analysis of airborne sulfur dioxide (SO₂). Samples are taken in the breathing zone of workplace personnel, and analysis is performed by ion chromatography (IC).

1.1. History

Previously, OSHA collected compliance samples for SO₂ exposure determinations in midjet-fritted glass bubblers containing 0.3 N (\approx 1%) hydrogen peroxide (H₂O₂) which converted the SO₂ to sulfuric acid (H₂SO₄). The amount of sulfate (SO₄²⁻) in the peroxide solution was measured by IC and gravimetrically converted to represent the amount of SO₂ collected (5.1.). Because bubblers are inconvenient to use as personal samplers due to spillage or breakage, it was desirable to develop a solid-sorbent sampling method. A method for collecting SO₂ was developed, which sampled SO₂ in the air using a sampling tube containing impregnated charcoal (5.2., 5.3.). The chemical(s) used for charcoal impregnation were not specified and appear to be proprietary; however, the reaction implies an oxidation of SO₂ using a metal hydroxide base. The impregnated charcoal oxidized SO₂ to SO₄²⁻, which was then desorbed using a weakly basic solution. An aliquot of the solution was analyzed by IC (5.2., 5.3.). Unfortunately, background levels of SO₄²⁻ found in the impregnated charcoal were considered unacceptable, especially when applying the current Permissible Exposure Limit (PEL) of 2 ppm. Previously, the OSHA Time Weighted Average (TWA) PEL was 5 ppm SO₂. Because of this contamination and the PEL reduction, OSHA reinstated use of the bubbler method listed in reference 5.1. until better methodology could be found.

Using the principle applied for the impregnated charcoal collection of SO₂, a new material, impregnated activated beaded carbon (IABC) was developed. The IABC has a significantly lower background level of SO₄²⁻ (< 3 μ g). This current method was evaluated using the IABC as the collection media.

1.2. Principle

Sulfur dioxide is collected using IABC sorbent which is contained in a glass tube. The collected SO₂ is converted to sulfite (SO₃²⁻) by the sorbent and then slowly oxidized to SO₄²⁻. This oxidation is augmented at the laboratory by addition of a desorbing solution containing 0.3 N (\approx 1%) H₂O₂ in 15 mM sodium hydroxide (NaOH) to each IABC sample. The resultant SO₄²⁻ is analyzed by IC using a conductivity detector; a gravimetric conversion is used to calculate the amount of SO₂ collected.

1.3. Advantages and Disadvantages

- 1.3.1. This method has adequate sensitivity for determining compliance with the OSHA Short-Term Exposure Limit (STEL) of 5 ppm and the TWA-PEL of 2 ppm for workplace exposures to SO₂.
- 1.3.2. The method is simple, rapid, and easily automated.
- 1.3.3. The SO₄²⁻ contaminant (background) levels of the IABC sorbent are very low (< 3 μ g), especially when compared to the impregnated charcoal previously used in OSHA Method No. ID-107.
- 1.3.4. A disadvantage is the need for a desorption efficiency (DE) correction which is mass-dependent and may be lot-dependent (also see Sections 3.7.3. and 4.1.).

1.4. Method Performance

A synopsis of method performance is presented below. Further information can be found in Section 4.

- 1.4.1. This method was validated over the concentration range of 1.36 to 4.16 ppm. An air volume of 12 L and a flow rate of 0.1 L/min were used.
- 1.4.2. The qualitative detection limit was 0.0187 μ g/mL or 0.187 μ g (as SO₄²⁻) when using a 10-mL solution volume. This corresponds to 0.005 ppm SO₂ for a 12-L air volume.
- 1.4.3. The quantitative detection limit was 0.0624 μ g/mL or 0.624 μ g (as SO₄²⁻) when using a 10-mL solution volume. This corresponds to 0.016 ppm SO₂ for a 12-L air volume. A 50- μ L sample

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loop and a detector setting of 1 microsiemens (μS) full-scale output were used.

- 1.4.4. The sensitivity of the analytical method, when using the instrumental parameters listed in Section 3.6., was calculated from the slope of a linear working range curve (0.5 to 10 $\mu\text{g}/\text{mL}$ SO_{42-}). The sensitivity was 2.2×10^7 area units per 1 $\mu\text{g}/\text{mL}$. A Dionex Series 4500i ion chromatograph with AI450 computer software was used (Dionex, Sunnyvale, CA).
- 1.4.5. This method compared favorably to OSHA Method no. ID-104 (modified) for SO_2 (5.1.) which served as the reference method.
- 1.4.6. A desorption efficiency (DE) correction is required at mass loadings up to 400 μg SO_2 (see Sections 3.7.3. and 4.1.).
- 1.4.7. The total pooled coefficient of variation (CV_T) for samples taken at about 0.5, 1, and 2 times the OSHA PEL (1 to 4 ppm for TWA-type samples) was 0.048. The method exhibited slight negative bias (-3.3%) for this concentration range after DE corrections were applied. Other concentration range and TWA results are shown below:

| | TWA | STEL | LOW |
|------|--------------|-------------|--------------|
| CV | 0.048 | 0.028 | 0.032 |
| Bias | -3.3% | +0.6% | -6.5% |
| OE | $\pm 12.9\%$ | $\pm 6.2\%$ | $\pm 12.9\%$ |

For the STEL evaluation, 5.32 ppm was used for test atmospheres. For sampling at a concentration (LOW) near what may be expected in indoor air monitoring, approximately 0.3 ppm SO_2 was used. Bias and overall error (OE) values were calculated from those found analytically versus theoretical (known) values. The theoretical concentrations were calculated from flows of a certified cylinder of SO_2 and dilution air.

- 1.4.8. The collection efficiency at 2 times the PEL was 100%. Samples were collected from a generated test atmosphere of 4 ppm SO_2 for 120 min.
 - 1.4.9. Breakthrough tests were performed at concentrations of 7.20 and 14.8 ppm SO_2 . No breakthrough was found for a sampling time of 240 min and an average sample flow rate of 0.1 L/min.
 - 1.4.10. Samples can be stored at ambient (20 to 25 $^\circ\text{C}$) temperature for a period of at least 30 days. Storage stability results show the mean sample recovery after 30 days was within $\pm 10\%$ of the theoretical calculations. Samples were stored on a laboratory bench.
- 1.5. Interferences
- 1.5.1. Other particulate sulfate compounds and H_2SO_4 will interfere in the analysis of SO_2 if they are collected in the IABC. Particulate and H_2SO_4 mist can be removed from the air during sampling using a modified sampling device which contains a Teflon[®] pre-filter (see Section 2.1.).
 - 1.5.2. Sulfur trioxide gas (SO_3), if present in a dry atmosphere, can give a positive bias in the SO_2 determination.
 - 1.5.3. Any substance that has the same retention time as SO_{42-} , when using the ion chromatographic operating conditions described in this method, is an interference. If the possibility of an interference exists, changing the separation conditions (column, eluent flow rate and strength, etc.) may circumvent the problem.

1.6. Source of Exposure

Sulfur dioxide is generated as a by-product from many industrial processes. These include the smelting of sulfide ores, the combustion of coal or fuel oils containing sulfur as an impurity, paper manufacturing, and petroleum refining (5.4.).

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1.7. Physical and Chemical Properties (5.4., 5.5.)

Sulfur dioxide (CAS No. 7446-09-5)

| | |
|------------------|-----------------|
| Chemical formula | SO ₂ |
| Formula weight | 64.07 |
| Melting point | -72.7 °C |
| Boiling point | -10.0 °C |
| Vapor density | 2.3 (air = 1) |

SO₂ is a colorless, nonflammable gas with a characteristic, strong, and suffocating odor. It is soluble in water, methanol, ethanol, chloroform, ethyl ether, acetic acid, and sulfuric acid.

1.8. Toxicology (5.6.)

Information listed within this section is a synopsis of current knowledge of the physiological effects of SO₂ and is not intended to be used as a basis for OSHA policy.

Sulfur dioxide is intensely irritating to the eyes and respiratory tract. Workplace exposure to SO₂ can cause both chronic and acute effects. The chronic effects of exposure include permanent pulmonary impairment, which is caused by repeated episodes of bronchoconstriction. It has been reported that workers exposure to high concentrations of SO₂ (80 to 100 ppm) may cause an increased incidence of nasopharyngitis, shortness of breath on exertion (dyspnea), and chronic fatigue. Concentrations of SO₂ from 2 to 36 ppm produced a significantly higher frequency of respiratory disease symptoms, including chronic coughing, expectoration, and dyspnea.

The acute effects include upper respiratory tract irritation, rhinorrhea, choking, and coughing. Within 5 to 15 minutes from the onset of exposure, workers develop temporary reflex bronchoconstriction and increased airway resistance.

2. Sampling

2.1. Equipment

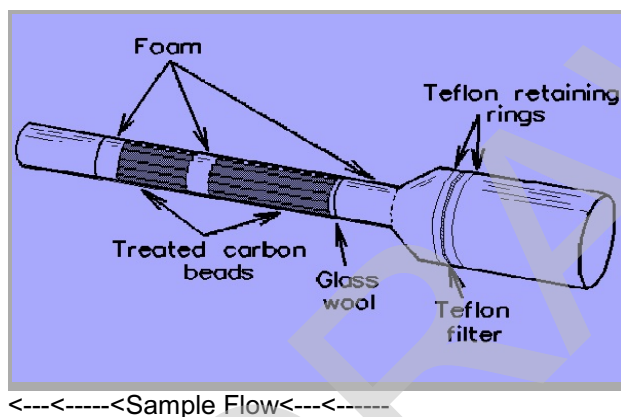
- 2.1.1 Calibrated personal sampling pumps capable of sampling within ±5% of the recommended flow rate of 0.1 L/min are used.
- 2.1.2. Solid sorbent sampling tubes are prepared using glass tubes, glass wool plugs, and IABC. Sampling tubes can be commercially obtained. Two types of sampling tubes are commercially available:

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Type I is a glass tube packed with a 100-mg IABC front and 50-mg backup section (Cat. No. 226-80, SKC Inc., Eighty Four, PA). The IABC is held in place with glass wool, foam, and a stainless steel retainer clip. If interference from particulate is probable, a prefilter/cassette sampling assembly can be used with this tube. See Section 2.1.5. for more details regarding the prefilter.

Type II, a combination sampling device (Forest Biomedical, Salt Lake City, UT) can be used to remove particulate and collect H_2SO_4 mist during SO_2 sampling. The combination device, as shown below, consists of two different glass tubes connected together. The front part of the tube contains a Teflon® filter, retaining rings, foam, and a glass wool plug. The Teflon® filter is used to trap any particulate and H_2SO_4 . The dimensions of the front portion of the sampling device are 12-mm o.d., 10-mm i.d., and 25-mm long. The second part of the device contains two sections of IABC and is used for collecting SO_2 . The dimensions of the second part are 6-mm o.d., 4-mm i.d., and 50-mm long. Both ends of the sampling tube are sealed with plastic caps.



Type II - Combination Sampling Device

If commercial tubes are unavailable, sampling tubes can be prepared using carbon bead impregnated in the same fashion as discussed in OSHA method ID-180 for phosphine (5.7.).

Note: The grade of carbon bead appears less significant for SO_2 when compared to phosphine (5.7.).

Prepare each tube for SO_2 collection with a 100-mg IABC front and a 50-mg backup section. Separate each IABC section using a small amount of glass wool.

2.1.3. A stopwatch and bubble tube or meter are used to calibrate pumps.

2.1.4. Various lengths of polyvinyl chloride tubing are used to connect sampling tubes to pumps.

2.1.5. If the workplace air being sampled is suspected of containing particulate which could interfere (i.e. H_2SO_4 or sulfates), the prefilter/cassette assembly listed below or a Type II sampling tube should be used.

- a) Filter for particulate collection, Teflon® (PTFE), 0.45 μm pore size, 25-mm diameter (part no. 130620, Nucleopore Corp., Pleasanton, CA)
- b) Carbon-filled polypropylene cassette, 25-mm diameter, (part no. 300075, Nucleopore) (See Section 4.10. for further details regarding this cassette)
- c) Porous plastic support pad (part no. 220600, Nucleopore)

Note: Do not use glass fiber prefilters for particulate collection during sampling for SO_2 . Loss of SO_2 can occur due to the slightly basic properties of these filters. See Section 4.10. for further details.

Assemble the prefilter assembly such that sampled air enters the Teflon® filter first and the

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plastic support pad faces the sampling tube. Use a minimum amount of tubing to connect the Type I sampling tube to the prefilter assembly.

2.2. Sampling Procedure (Bulk or wipe samples are not applicable)

- 2.2.1. Connect the sampling tube (Type I or II) to the calibrated sampling pump, making sure sampled air enters the large section (100 mg) of IABC first. Place the sampling device on the employee such that air is sampled from the breathing zone.
- 2.2.2. For STEL samples, use a flow rate of 0.1 L/min and a minimum sampling time of 15 min. For TWA determinations, take consecutive 12-L samples at a flow rate of 0.1 L/min for 120 min each. If possible, take enough consecutive samples to cover the entire work shift.
- 2.2.3. After sampling, place plastic end caps tightly on both ends of the tube and apply OSHA Form 21 seals. Record the sampling conditions.
- 2.2.4. Use the same lot of IABC tubes for blank and collected samples. Handle the blank sorbent tube in exactly the same manner as the sample tubes except that no air is drawn through it. Submit at least one blank tube for each batch of ten samples.
- 2.2.5. When other compounds are known or suspected to be present in the air, such information should be transmitted with the sample.
- 2.2.6. Specify SO₂ analysis and ship samples to the laboratory. If necessary, any Teflon® pre-filters used can be analyzed for H₂SO₄ (when H₂SO₄ is suspected to be present in the workplace atmosphere).

3. Analysis

3.1. Safety Precautions

- 3.1.1. Refer to appropriate IC instrument manuals and the Standard Operating Procedure (SOP) for proper instrument operation (5.8.).
- 3.1.2. Observe laboratory safety regulations and practices.
- 3.1.3. Sulfuric acid, sodium hydroxide, and hydrogen peroxide are corrosive. Use appropriate personal protective equipment such as safety glasses, gloves, and lab coat when handling corrosive chemicals. Prepare solutions in an exhaust hood.

3.2. Equipment

- 3.2.1. Ion chromatograph (Model 4000i or 4500i Dionex, Sunnyvale, CA) equipped with a conductivity detector.
- 3.2.2. Automatic sampler (Dionex Model AS-1) and sample vials (0.5 mL).
- 3.2.3. Laboratory automation system: Ion chromatograph interfaced with a data reduction system (AI450, Dionex).
- 3.2.4. Micromembrane suppressor, anion (Model AMMS-1, Dionex).
- 3.2.5. Separator and guard columns, anion (Model HPIC-AS4A and AG4A, Dionex).
- 3.2.6. Disposable syringes (1 mL).
- 3.2.7. Syringe pre-filters, 0.5- μ m pore size (part no. SLSR 025 NS, Millipore Corp., Bedford, MA).

Note: Some syringe pre-filters are not cation- or anion-free. Tests should be performed with blank solutions first to determine contamination and suitability with the analyte.

- 3.2.8. Miscellaneous volumetric glassware: Micropipettes, volumetric flasks, Erlenmeyer flasks, graduated cylinders, and beakers.

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- 3.2.9. Scintillation vials, glass, 20-mL.
- 3.2.10. Equipment for eluent degassing (vacuum pump, ultrasonic bath).
- 3.2.11. Analytical balance (0.01 mg).
- 3.3. Reagents - All chemicals should be at least reagent grade.

3.3.1. Principal reagents:

CAUTION:

NaOH, H₂SO₄, or 30% H₂O₂ can cause skin irritation or burns.

Sodium carbonate (Na₂CO₃)
Sodium bicarbonate (NaHCO₃)
Sodium hydroxide (NaOH)
Sulfuric acid (H₂SO₄), concentrated, 98%
Hydrogen peroxide (H₂O₂), 30%
Sodium sulfate (Na₂SO₄), anhydrous
Deionized water (DI H₂O) with a conductance of <10 µS.

- 3.3.2. Eluent (1.0 mM Na₂CO₃ + 1.0 mM NaHCO₃):
Dissolve 0.212 g Na₂CO₃ and 0.168 g NaHCO₃ in 2.0 L DI H₂O. Sonicate this solution and degas under vacuum for 15 min.
- 3.3.3. Suppressor regenerant solution (0.02 N H₂SO₄):
Carefully transfer 1.14 mL concentrated H₂SO₄ into a 2-L volumetric flask which contains approximately 500 mL DI H₂O. Dilute to volume with DI H₂O.
- 3.3.4. Desorbing solution [0.3 N (≈1%) H₂O₂ in 15 mM NaOH]:
Dissolve ≈0.6 g NaOH in approximately 500 mL of DI H₂O contained in a 1-L volumetric flask. Carefully add 34 mL of 30% H₂O₂ and then dilute to the 1-L mark with DI H₂O. Prepare weekly.
- 3.3.5. Sulfate (SO₄₂₋) stock standard (1,000 µg/mL):
Dissolve and dilute 1.4792 g of Na₂SO₄ to 1.0 L with DI H₂O. Prepare yearly.
- 3.3.6. Sulfate (SO₄₂₋) standard solutions, 100, 10, and 1 µg/mL:
Pipette appropriate volumes of the 1,000 µg/mL SO₄₂₋ stock standard into volumetric flasks and dilute to the mark with eluent. Prepare monthly.
- 3.4. Working Standard Preparation
 - 3.4.1. Prepare SO₄₂₋ working standards in eluent. A method for preparing a series of working standards using 10-mL final solution volumes is shown below:

| Working Std µg/mL | Std Solution (µg/mL) | Aliquot (mL) | Eluent Added (mL) |
|----------------------|-------------------------|-----------------|----------------------|
| 0.5 | 1 | 5 | 5 |
| 1 | 1 | * | * |
| 2 | 10 | 2 | 8 |
| 5 | 10 | 5 | 5 |
| 10 | 10 | * | * |
| 20 | 100 | 2 | 8 |
| 30 | 100 | 3 | 7 |
| 50 | 100 | 5 | 5 |

* Already prepared in Section 3.3.6.

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- 3.4.2. To prepare each working standard listed above, pipette an appropriate aliquot of the specified standard solution (prepared in Section 3.3.6.) and add the specified amount of eluent.
- 3.4.3. As an alternative, pipette each aliquot into a 10-mL volumetric flask and dilute to volume with eluent.

3.5. Sample Preparation

Note: If H₂SO₄ is a requested analyte and a Type II sampling device or a PTFE prefilter was used, see OSHA Stopgap Method ID-165SG or OSHA Method No. ID-113 for further details regarding sample preparation, analysis, and calculation of results for H₂SO₄.

- 3.5.1. Carefully remove and discard the rear glass wool plug (or foam for the Type II sampler) without losing any beaded carbon.

Note: The sorbent should always be removed from the glass tube via the opposite end of collection (i.e. 50-mg IABC backup section is removed first). This will minimize the possibility of contamination from any collected particulate.

- 3.5.2. Carefully transfer each IABC section from a sample tube and place in separate 25-mL Erlenmeyer flasks or scintillation vials.
- 3.5.3. Pipette 10 mL of desorbing solution into each flask. Cap each flask tightly and allow each solution to sit for at least 60 min. Occasionally swirl each solution.

3.6. Analysis

- 3.6.1. Pipette a 0.5- to 0.6-mL portion of each standard or sample solution into separate automatic sampler vials. Place a filtercap into each vial. The large filter portion of the cap should face the solution.
- 3.6.2. Load the automatic sampler with labeled samples, standards, and blanks.
- 3.6.3. Set up the ion chromatograph in accordance with the SOP (5.8.).

Note: An SOP is a written procedure for a specific instrument. It is suggested that SOPs be prepared for each type of instrument used in a lab to enhance safe and effective operation.

Typical operating conditions for a Dionex 4000i or 4500i with a conductivity detector and an automated sampler are listed below:

Ion Chromatograph

| | |
|----------------------------|---|
| Eluent | 1.0 mM Na ₂ CO ₃ /1.0 mM NaHCO ₃ |
| Column temperature: | ambient |
| Anion precolumn: | AG4A |
| Anion separator column: | AS4A |
| Anion suppressor: | AMMS-1 |
| Conductivity Output range: | 1 µS |
| Sample injection loop: | 50 µL |

Pump

| | |
|----------------|----------|
| Pump pressure: | ~900 psi |
| Flow rate: | 2 mL/min |

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Chromatogram

Run time: 10 min
Peak retention time: ~6 min for SO₄₂₋

3.6.4. Follow the SOP for further instructions regarding analysis (5.8.).

3.7. Calculations

3.7.1. After the analysis is completed, retrieve the peak areas or heights. Obtain hard copies of chromatograms from a printer.

3.7.2. Prepare a concentration-response curve by plotting the peak areas or peak heights versus the concentration of the SO₄₂₋ standards in µg/mL.

3.7.3. Perform a blank correction for each IABC front and backup sections. Subtract the µg/mL SO₄₂₋ blank value (if any) from each sample reading if blank and sample solution volumes are the same. If a different solution volume is used, subtract the total µg blank value from total µg sample values.

3.7.4. Calculate the air concentration of SO₂ (in ppm) for each air sample:

$$A = (\mu\text{g/mL SO}_4^{2-}) \times (\text{Sol Vol}) \times (\text{GF})$$

$$\text{corr } \mu\text{g SO}_2 = \frac{A}{DE}$$

$$\text{ppm SO}_2 = \frac{(\text{corr } \mu\text{g SO}_2) \times (\text{Mol Vol})}{(\text{AV}) \times (\text{Mol Wt})}$$

Where:

A = uncorrected µg SO₂
corr µg SO₂ = µg SO₂ with DE correction applied
µg/mL SO₄₂₋ = Amount found (from calibration curve)
Sol Vol = Solution volume (mL) from Section 3.5.3.
GF, SO₂/SO₄₂₋ = Gravimetric factor = 0.667
Mol Vol = Molar volume (L/mol) = 24.45 (25 °C and 760 mmHg)
AV = Air volume (L)
Mol Wt = Molecular weight for SO₂ = 64.0 (g/mol)
DE = Desorption efficiency (mass-dependent, see sliding scale below or equation in 3.7.5.)

Amt of SO₂ (µg) DE Factor

| | |
|------------|-------|
| ≤ 30 | 0.800 |
| 31 to 50 | 0.825 |
| 51 to 75 | 0.850 |
| 76 to 100 | 0.875 |
| 101 to 200 | 0.900 |
| 201 to 400 | 0.950 |
| >400 | 1.000 |

3.7.5. An alternative to the DE correction sliding scale above is the following equation:

$$DE = -1.1386 \times 10^{-6} (A)^2 + 1.0037 \times 10^{-3} (A) + 7.81 \times 10^{-1}$$

Where: A = uncorrected µg SO₂

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3.7.6. The DE correction may be lot-dependent. The corrections stated in this method were determined using lot no. 673 (Cat. no. 226-80, SKC Inc., Eighty Four, PA) or beaded carbon prepared for sampling phosphine (5.7.). A difference in DE was not noted for these two preparations (Section 4.1.2.). Future lots or different grades should be evaluated for DE corrections.

3.8. Reporting Results

Add the backup section ppm SO₂ result (if any) to the front section result for each sample. Report results to the industrial hygienist as ppm SO₂.

4. Backup Data

This method has been validated for a 12-L, 120-min sample taken at a flow rate of 0.1 L/min. The method validation was conducted near the OSHA TWA-PEL and STEL of 2 ppm and 5 ppm, respectively. The sampling tubes used during the validation consisted of a two-section tube (Type I) packed with a 100-mg IABC front and 50-mg backup section or Type II (see Section 2.1.2). Tubes were obtained commercially (Lot no. 673, Cat. no. 226-80, SKC Inc., Eighty Four, PA) or prepared in-house. Preliminary tests were conducted using a 10% (w/w) impregnation of a metal hydroxide base on the carbon bead; however, tests conducted at high humidity (80%) indicated a formation of a slurry inside the sampling tube. The amount of metal hydroxide base was lowered to 1% during validation. A difference in SO₂ results using either 1 or 10% base impregnation was not noted; an exception was the 80% RH tests where slurry formation made it difficult to remove the carbon bead from the glass tubes.

The validation consisted of the following experiments:

1. An analysis of 19 samples (6 samples each at 2 × and 1 × TWA-PEL, and 7 samples at 0.5 × TWA-PEL) for the DE study.
2. A sampling and analysis of 16 samples (6 samples each at 2 × and 1 × TWA-PEL, and 4 samples at 0.5 × TWA-PEL) collected from dynamically generated test atmospheres at 50% RH. Samples at a concentration near the STEL and at concentrations expected during indoor air quality investigations (≈0.3 ppm) were also taken.
3. A determination of the sampling media collection efficiency at approximately 4 ppm (2 × TWA-PEL).
4. A determination of breakthrough.
5. An evaluation of storage stability at 20 to 25 °C for 24 collected samples.
6. A determination of any significant effects on results when sampling at different humidities.
7. A determination of the qualitative and quantitative detection limits.
8. A comparison of methods.
9. Evaluation of the Type II sampling tube for collecting SO₂.
10. Evaluation of a prefilter/cassette assembly for use with Type I samplers.
11. Summary.

All theoretical (known) concentrations of generated test atmospheres were calculated from controlled flows of a cylinder of 303 ppm SO₂ in nitrogen (certified concn, Alphagaz, LaPorte, TX) and dilution air. An analysis of the cylinder concentration using OSHA Method No. ID-104 (modified) (5.1.) indicated the manufacturer's stated concentration was accurate. The OSHA method ID-104 was modified such that the 0.3 N H₂O₂ sample collection solution was made more basic by addition of NaOH. The final collection solution used for the modified method ID-104 was 0.3 N H₂O₂ in 15 mM NaOH.

A generation system was assembled, as shown in Figure 1, and used for all experiments except detection limit determinations. Samples using OSHA Method ID-104 (modified) were taken side-by-side with any IABC samples. All samples were analyzed by IC. The IABC samples were subject to the sliding scale DE corrections listed in Section 3.7.3 (with the obvious exception of those samples used to calculate DE corrections in Section 4.1.). A correction was not applied nor necessary for the liquid sorbent sampler results (ID-104 modified), or sulfate solutions used to calculate detection limits.

All results were calculated from concentration-response curves and statistically examined for outliers. In addition, the analysis (Section 4.1.) and sampling and analysis results (Section 4.2.) were tested for homogeneity of variance. Possible outliers were determined using the Treatment of Outliers test (5.9.). Homogeneity of variance was determined using the Bartlett's test (5.10.). Statistical evaluation was conducted according to Inorganic Methods Evaluation Protocol (5.11.). The overall error (OE) (5.11.) was calculated using the equation:

$$OE_i = \pm(|\text{bias}_i| + 2CV_i) \times 100\% \text{ (95\% confidence level)}$$

Where i is the respective sample pool being examined.

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4.1. Analysis

Nineteen samples were prepared by adding known amounts of SO₂ to the IABC tubes to determine recoveries (DE) for the analytical portion of the method. For this experiment, an active method of spiking with low-flow (0.03 L/min) sampling pumps was used to determine the amount of gas collected and not necessarily the sampling capability of the IABC at the low-flow rate.

4.1.1. Procedure: Sampling tubes containing IABC were spiked using the low flow pumps and generation system as described in references 5.7. and 5.12. The SO₂ source mentioned previously was diluted to approximately 30 ppm using the generation system shown in Figure 1. Calibrated low-flow-rate pumps were connected to the sampling manifold and were used to deliver the spikes for measured time periods. Air used to dilute the SO₂ source was tempered to 50% RH and 25 °C. Pumps used for this experiment (Miniature Personal Air Sampling Pumps, Model No. 222-3-12, SKC Inc., Eighty Four, PA) were calibrated to collect samples at 0.030 L/min. Spikes were approximately 32, 64, 130, and 400 µg SO₂. These levels correspond approximately to 0.5, 1, 2, and 6 times the PEL for a 12-L air sample at a 0.1-L/min flow rate.

4.1.2. Results: Desorption efficiencies, presented in Table 1, varied depending on the amount of SO₂ collected. For 0.5 times the PEL (32 µg SO₂), the DE was close to 0.8; the DE was 1.0 for 6 × PEL (≈400 µg SO₂). The DE corrections are similar (although larger) to those found during a previous validation of a sorbent tube (Method no. ID-107, see reference 5.3.) for SO₂ collection. The previous method used charcoal impregnated in a fashion similar to the IABC sorbent. The average DE correction for the lot of treated charcoal specified in Method no. ID-107 was 0.927, and resulted from averaging DEs of 0.909 (168 µg SO₂), 0.934 (333 µg SO₂), and 0.940 (669 µg SO₂).

A mechanism to explain the decrease in DE at low mass loadings has not been found; however, it is possible that a portion of the initial SO₂ entering the sampling tube strongly bonds with the surface of the carbon bead matrix and becomes unrecoverable. A strong and weak bonding mechanism for SO₂ has been noted on active carbon surfaces (5.13.). Once the strong bonding sites are occupied, a DE closer to what is expected is achieved. At larger mass loadings the amount of strongly bonded SO₂ would remain the same, and become insignificant when compared to the total amount collected; thus the DE approaches unity.

A test of two different grades of beaded carbon (impregnated) was also conducted and a significant difference in DE was not noted:

(Sampling Time = 120 min, flow rate ≈ 0.1 L/min, SO₂ concentration = 1.19 ppm, 25 °C, 50% RH)

| Grade | SKC Bulk | SKC lot no. 745* |
|--------------------------------|----------|------------------|
| Sample Type | General | MU-AZ |
| N | 3 | 3 |
| Mean (ppm SO ₂) | 1.19** | 1.20** |
| Std Dev (ppm SO ₂) | 0.015 | 0.006 |
| CV | 0.013 | 0.005 |
| Recovery | 100% | 101% |

* SKC Catalog no. 226-32 (These tubes were manufactured for the collection of phosphine. The appropriate amount of IABC was removed from a tube, placed in a Type II sampling device, and used to collect SO₂).

** DE corrected using scale shown in Section 3.7.3.

Grades listed above are assigned by the manufacturer of the untreated beaded carbon (Kureha Chemical, NY) to designate bead type and manufacturing process used. The general grade is a more common bead which was found to have poor retention efficiency for collection of phosphine (5.7.).

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Although the data shows comparable DEs, even for different grades, future lots or grades of beaded carbon should be evaluated for DE corrections.

4.2. Sampling and Analysis

To determine the precision and accuracy of the method, known concentrations of SO₂ were generated, samples were collected, prepared, and analyzed.

4.2.1. Procedure:

- 1) The SO₂ source mentioned previously was used to generate test atmospheres of SO₂. This source was diluted with filtered, humidified air using the system shown in Figure 1.
- 2) Dynamic generation system
A Miller-Nelson Research Inc. flow, temperature, and humidity control system (Model HCS-301, Monterey, CA) was used for air flow control and conditioning. All generation system fittings and connections were Teflon®. A glass mixing chamber was used to mix the tempered, filtered air with the contaminant gas. The system was set to generate test atmospheres at 50% RH and 25 °C.
- 3) The SO₂ and diluent air flow rates were adjusted using mass flow controllers. Flow rates were also measured using a dry test meter (for diluent air) and a soap bubble flow meter (for SO₂ gas).
- 4) Samples were taken from the sampling manifold using constant-flow pumps. Calibrated P125 and Alpha 2 pumps (E.I. duPont de Nemours & Co., Wilmington, DE) were used. Pump flow rates were approximately 0.1 L/min and sampling time was 120 min. Sample concentrations were approximately 0.5, 1, and 2 times the OSHA TWA-PEL for 12-L air samples. For samples taken near the STEL (~5.8 ppm SO₂ was used), a 0.1 L/min sampling rate for 15 min was used. For low concentration samples, a test atmosphere of approximately 0.3 ppm was used.

4.2.2. Results: The results for TWA, STEL, and low concentration-type exposures are shown in Tables 2 and 3a-3b, respectively. The test atmosphere sample (Table 2) and spiked sample (Table 1) results for TWA samples passed the Bartlett's test and were pooled to determine a total CV (CV_T) for the sampling and analytical method. For the experiments, the pooled coefficients of variation, bias, and OE are as follows:

| | TWA | STEL | LOW |
|------|--------|---------|---------|
| CV | 0.048* | 0.028** | 0.032** |
| Bias | -3.3% | +0.6% | -6.5% |
| OE | ±12.9% | ±6.2% | ±12.9% |

* CV_T(pooled), see reference 5.11. for further details. The CV₁ is taken from 0.5, 1, and 2 × PEL results. The 6 × PEL level is not included in statistical calculations.

**CV₂ only

4.3. Collection Efficiency

Procedure: Six commercially-prepared (Type I) sampling tubes were used for collection at a concentration of approximately 2 times the OSHA TWA-PEL for 120 min at 0.1 L/min (50% RH and 25 °C). The amounts of SO₂ vapor collected in the first section (100 mg of sorbent) and second section (50 mg of sorbent) were determined. The collection efficiency (CE) was calculated by dividing the amount of SO₂ collected in the first section by the total amount of SO₂ collected in the first and second sections.

Results: The results in Table 4 show a CE of 100%. No SO₂ was found in the second sorbent

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section for the CE experiment.

4.4. Breakthrough Study

(Note: Breakthrough is defined as >5% loss of analyte through the sampling media at 50% RH)
Procedure: Two separate experiments were conducted to test for breakthrough. For the first experiment, the same procedure as the CE test was used with two exceptions:

- 1) The concentration was increased to a level approximately 4 times the TWA-PEL (7.20 ppm SO₂).
- 2) Samples were collected at 0.1 L/min for 240 min.

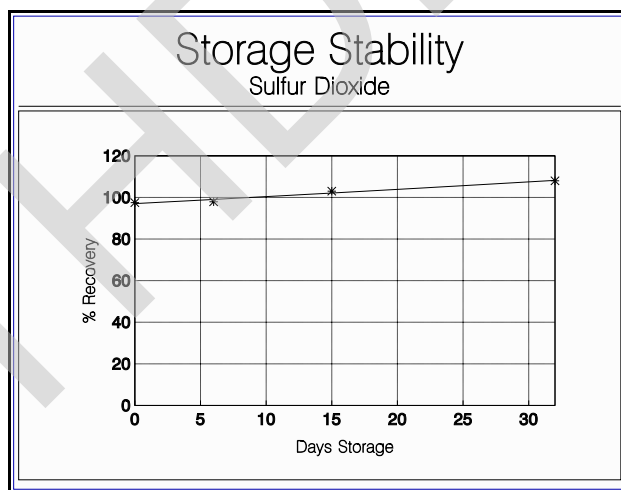
The second experiment was conducted using the Type II sampling tube (Section 2.1.2.) at a concentration larger than expected during routine sampling of industrial hygiene operations (14.8 ppm SO₂). Experimental parameters for this test were: Four Type II sampling tubes, 25 °C, 50% RH, 240-min sampling time, 0.1 L/min sample rate, and 14.8 ppm SO₂ test concentration.

The amount of breakthrough for each sampling tube was calculated by dividing the amount collected in the second section by the total amount of SO₂ collected in the first and second sections.

Results: No breakthrough of SO₂ into the second section was found. Results for both concentrations are shown in Table 5.

4.5. Storage Stability

Procedure: A study was conducted to assess the storage stability at 20 to 25 °C of the sampling media after SO₂ collection. Twenty-four samples were taken near the OSHA TWA-PEL of 2 ppm. After collection, all samples were stored under normal laboratory conditions (20 to 25 °C) on a lab bench and were not protected from light. Six samples were initially desorbed and analyzed, then six samples were desorbed and analyzed after various periods of storage (6, 15, and 32 days).
Results: The mean of samples analyzed after 32 days was within 10% of the theoretical value as shown in Table 6. A slight increase in recovery over time was noted as shown in the figure below.



4.6. Humidity Study

Note: Prior to the validation, preliminary humidity tests using a beaded carbon impregnated with a 10% (w/w) load of a metal hydroxide base indicated acceptable SO₂ collection at 25 and 50% RH (assuming 25 °C). Although data was also acceptable at 80% RH (2-h sampling time), a slurry developed inside the sampling tube. The resulting slurry presented some difficulty in removing the bead from the sampling tube and required an enhanced attention to technique. The slurry appeared to be a combined result of the following factors:

- 1) the hydrophilic nature of the impregnated base,

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2) the hydrophobicity of the beaded carbon, and
3) complications arising from limited temperature control during sample generations at 80% RH.

Factor (1) appeared to be the main factor in producing a slurry. The "wetting" effect apparently was assisted by differences in temperature during generation. The high humidity tests were conducted during the winter season and the ambient temperature in the laboratory was about 18 °C. The ambient temperature is normally near the test atmosphere temperature of 25 °C; due to a malfunctioning room thermostat it was somewhat cooler during these tests. The temperature of the sampling tubes are ambient because of the sampling manifold design. The test atmosphere condensation inside the tube probably was accelerated when entering the cooler sampling tube.

The amount of base impregnation was lowered to 1% (w/w) to alleviate potential formation of a slurry.

Procedure: A study was conducted to determine any effect on results when samples are collected in different humidities. Samples were taken using the generation system and procedure described in Section 4.2. Test atmospheres were generated at 25 °C and at approximately 0.5, 1, and 2 times the OSHA TWA-PEL. Relative humidities of 30%, 50%, and 80% were used at each concentration level tested.

Results: Results of the humidity tests are listed in Table 7. An F test was used to determine if any significant effect occurred when sampling at different humidities. As shown, the calculated F values exceeded critical F values (5.14.) for all the concentrations tested and a significant difference in results occurred across the humidity ranges tested. An examination of the data indicates a decrease in recoveries as humidity increases; however, a correction for humidity effect was not instituted because results at higher humidities were considered acceptable in terms of overall error (< ±25%).

The finding that increasing humidity produces a decrease in the amount of sulfate found appears to contradict previous findings in the literature (5.15.). The previous study (5.15.) indicated the conversion of SO₂ to sulfate on an active carbon surface was facilitated by the presence of an aqueous environment; however, testing carbons impregnated with a metal hydroxide base was not performed. The base appears to significantly alter the adsorption characteristics of the beaded carbon for SO₂.

4.7. Qualitative and Quantitative Detection Limit Study

Procedure: Low concentration samples were prepared by spiking desorbing solutions (Section 3.3.4.) with aliquots of aqueous standards prepared from sodium sulfate. These samples were analyzed using a 50-µL sample injection loop and a detector setting of 1 microsiemens (µS). A derivation of the International Union of Pure and Applied Chemistry (IUPAC) detection limit equation (5.16.) was used for this study.

Results: The results are shown in Table 8 for qualitative and quantitative detection limits, respectively. The qualitative limit is 0.0187 µg/mL as SO₄₂₋ at the 99.86% confidence level. The quantitative limit (99.99% confidence) is 0.0642 µg/mL as SO₄₂₋. Using a 12-L air volume and a 10-mL sample solution volume, the qualitative limit is 0.004 ppm and the quantitative limit is 0.013 ppm as SO₂.

4.8. Comparison of Methods

Procedure: In order to compare the performance of this method and to confirm the theoretical SO₂ concentrations, an independent method (OSHA Method no. ID-104 for SO₂, modified) was used. The collection solution for Method ID-104 was modified as previously mentioned in Section 4, and impingers were used to collect samples. The IABC and impinger samples were collected side-by-side from the generation system. All samples were analyzed by IC.

Results: Table 9 shows the results for different SO₂ concentrations. As shown, the theoretical concentration of the generation system, the IABC, and impinger results are in good agreement.

4.9. Type II Tube Study

Procedure: Eight Type II sampling tubes (see Section 2.1.2. for a description of this tube) were chosen for a statistical study using the generation system described in Section 4.2. These tubes contained PTFE membranes as prefilters. Samples were collected side-by-side with impinger

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samples at 25 °C, 80% RH, and at approximately the OSHA TWA-PEL for 120 min. The high humidity was selected as a worst-case test (Section 4.6.).

Results: Results are listed in Table 10. As shown, the Type II sampling device can be used to collect SO₂ without significantly altering the concentration. A slight increase in recovery was noted when compared to the earlier Type I tube results at 80% RH (Section 4.6.).

4.10. Prefilter Evaluation

Procedure: Past research regarding aerosols (5.17.) has indicated that particulate in the air sampled may penetrate any glass wool plugs and deposit on the sorbent when using conventional sampling tubes. To remedy this, a prefilter is generally used to stop the particulate before entry into the sampling tube. A preliminary experiment was conducted using the Type II sampling tube with glass fiber prefilters instead of PTFE. Significant losses were noted and were apparently caused by the slightly basic glass fiber filters reacting with some of the SO₂.

To further evaluate the possibility of SO₂ reacting with a prefilter/cassette sampling device, an experiment was performed using six Type I sampling tubes with prefilter sampling assemblies consisting of PTFE filter/polypropylene backup pad/carbon-filled cassettes. These cassettes (as specified in Section 2.1.5.) have enjoyed popularity for asbestos sampling (5.18.) and for their limited reactivity to certain corrosive gases (5.19.).

The test was conducted by taking six IABC samples without prefilters side-by-side with six IABC with prefilters. Samples were taken such that the test atmosphere entered the prefilter assembly first and then entered the IABC with minimal contact at the connection between cassette and sampling tubes. Small pieces of Tygon® tubing were used to connect the cassettes and IABC sampling tubes. All samples were taken at a flow rate of about 0.1 L/min for 120 min. The generation system concentration was approximately the TWA-PEL.

Results: The results of the comparison of IABC samples taken with and without prefilters is shown in Table 11. As shown, a difference in the amount of SO₂ collected was not noted between the prefilter/IABC and IABC sampling assembly. The PTFE prefilter/cassette assembly does not appear to inhibit the collection of SO₂ when using the stated sampling conditions.

4.11. Summary

The validation results indicate the method meets both the NIOSH and OSHA criteria for accuracy and precision (5.10., 5.11.). Collection efficiency, breakthrough, and storage stability are adequate. Although it appears that humidity effects are significant when sampling at different humidities, the results are within an acceptable range (OE < ±25%). Detection limits are adequate when samples are taken for 120 min at 0.1 L/min, or for 15-min STEL determinations. The method is adequate for monitoring TWA, STEL, and indoor air types of exposures.

The contaminant levels of blank IABC samples, when compared to levels found in the sorbent used in OSHA Method no. ID-107, were significantly lower. Blanks from IABC sorbent contained only 1 to 2 µg background (as SO₄₂₋), based on a sample solution volume of 10 mL. The previous impregnated charcoal sorbent contamination ranged from 20 to 100 µg SO₄₂₋.

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Note: OSHA no longer uses or supports this method (December 2019).

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Table 1
 Sulfur Dioxide Analysis - Desorption Efficiency (DE)

| Taken ($\mu\text{g SO}_2$) | Found ($\mu\text{g SO}_2$) | Recovery (F/T) | N | Mean DE | Std Dev | CV ₁ |
|---------------------------------|---------------------------------|-------------------|---|---------|---------|-----------------|
| 0.5×PEL | | | | | | |
| 31.850 | 25.330 | 0.795 | | | | |
| 32.090 | 24.280 | 0.757 | | | | |
| 32.090 | 26.730 | 0.833 | | | | |
| 32.090 | 28.290 | 0.882 | | | | |
| 32.090 | 24.930 | 0.777 | | | | |
| 32.090 | 25.530 | 0.796 | | | | |
| 32.090 | 25.610 | 0.798 | 7 | 0.805 | 0.041 | 0.051 |
| 1×PEL | | | | | | |
| 64.200 | 52.080 | 0.811 | | | | |
| 64.200 | 52.850 | 0.823 | | | | |
| 64.200 | 54.580 | 0.850 | | | | |
| 64.200 | 53.680 | 0.844 | | | | |
| 64.350 | 54.210 | 0.834 | | | | |
| 65.280 | 54.430 | 0.827 | 6 | 0.832 | 0.014 | 0.017 |
| 2×PEL | | | | | | |
| 131.340 | 113.960 | 0.868 | | | | |
| 131.340 | 116.950 | 0.890 | | | | |
| 131.340 | 111.350 | 0.848 | | | | |
| 131.340 | 116.130 | 0.884 | | | | |
| 131.340 | 120.030 | 0.914 | | | | |
| 128.400 | 115.780 | 0.902 | 6 | 0.884 | 0.024 | 0.027 |
| 6×PEL | | | | | | |
| 386.130 | 374.590 | 0.970 | | | | |
| 394.960 | 408.760 | 1.035 | | | | |
| 403.320 | 407088 | 1.011 | | | | |
| 450.430 | 380.990 | 0.846* | | | | |
| 409.050 | 395.910 | 0.968 | | | | |
| 435.440 | 4390.59 | 1.010 | 5 | 0.999 | 0.030 | 0.030 |

* Outlier, was deleted from final statistical calculations

F/T = Found/Taken DE = Desorption Efficiency

CV₁ (Pooled) = 0.036 (Calculated from pooling 0.5, 1, and 2 × PEL data only)

The mean DEs were significantly less than 1.0; therefore, a DE correction is needed. No corrections are necessary for SO₂ >400 μg .

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Table 2
 Sampling and Analysis - TWA Determination*

| Taken ($\mu\text{g SO}_2$) | Found ($\mu\text{g SO}_2$) | Recovery (F/T) | N | Mean | Std Dev | CV ₂ | OE |
|---------------------------------|---------------------------------|-------------------|---|-------|---------|-----------------|------|
| 0.5×PEL | | | | | | | |
| 1.360 | 1.340 | 0.985 | 4 | 0.991 | 0.042 | 0.043 | 9.2 |
| 1.360 | 1.420 | 1.044 | | | | | |
| 1.360 | 1.280 | 0.941 | | | | | |
| 1.360 | 1.350 | 0.993 | | | | | |
| 1×PEL | | | | | | | |
| 2.400 | 2.190 | 0.912 | 6 | 0.975 | 0.037 | 0.038 | 10.0 |
| 2.400 | 2.390 | 0.996 | | | | | |
| 2.400 | 2.400 | 1.000 | | | | | |
| 2.400 | 2.360 | 0.983 | | | | | |
| 2.400 | 2.42 | 1.008 | | | | | |
| 2.400 | 2.280 | 0.950 | | | | | |
| 2×PEL | | | | | | | |
| 4.160 | 4.040 | 0.971 | 6 | 0.944 | 0.051 | 0.054 | 16.5 |
| 4.160 | 3.690 | 0.887 | | | | | |
| 4.160 | 4.070 | 0.978 | | | | | |
| 4.160 | 3.870 | 0.930 | | | | | |
| 4.160 | 3.690 | 0.887 | | | | | |
| 4.160 | 4.210 | 1.012 | | | | | |

F/T = Found/Taken OE = Overall error (±%)
 Bias = -0.033
 CV₂ (Pooled) = 0.046
 CV_T (Pooled) = 0.048
 Overall Error (Total) = 12.9%
 * Samples were taken for 2 h.

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Table 3a
 Sampling and Analysis - STEL Determination

| Sample No. | Air Vol (L) | ppm SO ₂ | | N | Statistical Analysis | | | Recovery (%) |
|------------|-------------|---------------------|-------|---|----------------------|---------|-------|--------------|
| | | Found | Taken | | Mean | Std Dev | CV | |
| 1 | 1.33 | | | | | | | |
| 2 | 1.32 | 5.26 | 5.32 | | | | | |
| 3 | 1.50 | 5.35 | 5.32 | | | | | |
| 4 | 1.33 | 5.56 | 5.32 | | | | | |
| | | 5.22 | 5.32 | 4 | 5.35 | 0.15 | 0.028 | 100.6 |

Table 3b
 Sampling and Analysis - Low Concentration

| Sample No. | Air Vol (L) | ppm SO ₂ | | N | Statistical Analysis | | | Recovery (%) |
|------------|-------------|---------------------|-------|---|----------------------|---------|-------|--------------|
| | | Found | Taken | | Mean | Std Dev | CV | |
| 1 | 12.0 | 0.288 | 0.310 | | | | | |
| 2 | 12.6 | 0.288 | 0.310 | | | | | |
| 3 | 11.0 | 0.300 | 0.310 | | | | | |
| 4 | 12.0 | 0.288 | 0.310 | | | | | |
| 5 | 12.6 | 0.300 | 0.310 | | | | | |
| 6 | 11.3 | 0.275 | 0.310 | 6 | 0.290 | 0.009 | 0.032 | 93.5* |

*A DE correction of 0.8 was used from the scale listed in Section 3.7.4. Low mass loadings (<30 µg) may be more accurately corrected using the equation discussed in Section 3.7.5.

Table 4
 Collection Efficiency
 (2×TWA-PEL, 25 °C and 50% RH)

| Sample No. | ppm SO ₂ Found | | Collection Efficiency (%) |
|------------|---------------------------|----------------|---------------------------|
| | First Section | Second Section | |
| 1 | 4.04 | ND | 100.0 |
| 2 | 3.69 | ND | 100.0 |
| 3 | 4.07 | ND | 100.0 |
| 4 | 3.87 | ND | 100.0 |
| 5 | 3.69 | ND | 100.0 |
| 6 | 4.21 | ND | 100.0 |

Notes: (a) Sampled at 0.1 L/min for 120 min.
 (b) Samples were desorbed using a sample solution volume = 10.0 mL
 (c) ND = None detectable (<0.2 µg/mL as SO₄²⁻)

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Table 5
 Breakthrough Study

| ppm SO ₂ Found | | | |
|---------------------------|--------------|--------------|------------------|
| Sample No. | 1 st Section | 2 nd Section | Breakthrough (%) |
| 1 | 7.28 | ND | 0 |
| 2 | 6.09 | ND | 0 |
| 3 | 6.97 | ND | 0 |
| 4 | 7.27 | ND | 0 |
| 5 | 15.08 | ND | 0 |
| 6 | 15.10 | ND | 0 |
| 7 | 14.28 | ND | 0 |
| 8 | 13.64 | ND | 0 |

Notes: (a) Sampled at 0.1 L/min for 240 min
 (b) Samples were desorbed using a sample solution volume = 10.0 mL. A 10-fold dilution of sample solution was performed on samples 5-8.
 (c) ND = None detectable (<0.004 ppm SO₂)
 (d) Samples 1-4 used Type I sampling device; samples 5-8 used Type II

Table 6
 Storage Stability Test
 (1×TWA-PEL, 25 °C and 50% RH)

| (ppm SO ₂) | | | | | | | | |
|------------------------|-------------|-------|-------|---|------|---------|-------|--------------|
| Day | air Vol (L) | Found | Taken | N | Mean | Std Dev | CV | Recovery (%) |
| 0 | 11.5 | 2.19 | 2.40 | 6 | 2.34 | 0.088 | 0.038 | 97.5 |
| | 10.6 | 2.39 | 2.40 | | | | | |
| | 10.7 | 2.40 | 2.40 | | | | | |
| | 11.5 | 2.36 | 2.40 | | | | | |
| | 10.6 | 2.42 | 2.40 | | | | | |
| | 11.5 | 2.28 | 2.40 | | | | | |
| 6 | 11.9 | 2.21 | 2.39 | 6 | 2.43 | 0.130 | 0.054 | 103 |
| | 10.5 | 2.44 | 2.39 | | | | | |
| | 10.6 | 2.26 | 2.39 | | | | | |
| | 11.7 | 2.33 | 2.39 | | | | | |
| | 10.3 | 2.43 | 2.39 | | | | | |
| | 10.4 | 2.34 | 2.39 | | | | | |
| 15 | 11.8 | 2.34 | 2.35 | 6 | 2.53 | 0.068 | 0.027 | 108 |
| | 10.5 | 2.64 | 2.35 | | | | | |
| | 10.6 | 2.32 | 2.35 | | | | | |
| | 11.8 | 2.31 | 2.35 | | | | | |
| | 10.5 | 2.51 | 2.35 | | | | | |
| | 10.6 | 2.43 | 2.35 | | | | | |
| 32* | 8.2 | | 2.35 | 6 | 2.53 | 0.068 | 0.027 | 108 |
| | 7.9 | | 2.35 | | | | | |
| | 8.0 | | 2.35 | | | | | |
| | 8.2 | | 2.35 | | | | | |
| | 7.9 | | 2.35 | | | | | |
| | 8.0 | | 2.35 | | | | | |

* 90-min sampling time was used for this test only. The rest of the samples were taken for 120 min.

Note: OSHA no longer uses or supports this method (December 2019).

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Table 7
 Humidity Test

| | 30 | 50 | 80 |
|---------------------------|-------|-------|-------|
| RH (%) | 30 | 50 | 80 |
| ppm SO ₂ Taken | 0.990 | 1.48 | 1.19 |
| ppm SO ₂ Found | 1.08 | 1.33 | 1.02 |
| | 0.994 | 1.41 | 1.04 |
| | 0.998 | 1.27 | 1.05 |
| | 1.13 | 1.35 | 1.05 |
| | 1.05 | | 1.10 |
| | 1.11 | | 1.02 |
| N | 6 | 4 | 6 |
| Mean (ppm) | 1.06 | 1.34 | 1.05 |
| Std Dev (ppm) | 0.057 | 0.058 | 0.029 |
| CV | 0.054 | 0.043 | 0.028 |
| Ave Recovery (%) | 107 | 90.5 | 88.0 |

At the 99% confidence level:
 $F_{crit} = 6.70$ $F_{calc} = 33.81$ (2, 13 degrees of freedom)
 $F_{crit} < F_{calc}$; therefore, a significant difference in results was noted across the humidity levels tested.

Table 7a
 Humidity Test

| | 30 | 50 | 80 |
|---------------------------|-------|-------|-------|
| RH (%) | 30 | 50 | 80 |
| ppm SO ₂ Taken | 2.33 | 2.40 | 2.48 |
| ppm SO ₂ Found | 2.44 | 2.19 | 2.25 |
| | 2.50 | 2.39 | 2.26 |
| | 2.48 | 2.40 | 2.20 |
| | 2.48 | 2.36 | 2.40 |
| | 2.32 | 2.42 | 2.19 |
| | 2.51 | 2.28 | 2.36 |
| N | 6 | 6 | 6 |
| Mean (ppm) | 2.46 | 2.34 | 2.28 |
| Std Dev (ppm) | 0.070 | 0.088 | 0.085 |
| CV | 0.029 | 0.038 | 0.038 |
| Ave Recovery (%) | 105 | 97.5 | 91.8 |

At the 99% confidence level:
 $F_{crit} = 6.36$ $F_{calc} = 24.22$ (2, 15 degrees of freedom)
 $F_{crit} < F_{calc}$; therefore, a significant difference in results was noted across the humidity levels tested.

Table 7b
 Humidity Test

| | 30 | 50 | 80 |
|---------------------------|-------|-------|-------|
| RH (%) | 30 | 50 | 80 |
| ppm SO ₂ Taken | 4.44 | 4.40 | 4.40 |
| ppm SO ₂ Found | 4.28 | 4.04 | 4.03 |
| | 4.32 | 3.69 | 3.67 |
| | 4.25 | 4.07 | 3.94 |
| | 4.41 | 3.87 | 3.81 |
| | 4.72 | 3.57 | 3.96 |
| | | | 4.21 |
| N | 6 | 6 | 5 |
| Mean (ppm) | 4.43 | 3.91 | 3.88 |
| Std Dev (ppm) | 0.188 | 0.244 | 0.143 |
| CV | 0.042 | 0.062 | 0.037 |
| Ave Recovery (%) | 99.7 | 88.8 | 88.2 |

Note: OSHA no longer uses or supports this method (December 2019).

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At the 99% confidence level:

$F_{crit} = 6.52$ $F_{calc} = 11.88$ (2, 14 degrees of freedom)

$F_{crit} < F_{calc}$; therefore, a significant difference in results was noted across the humidity levels tested.

Table 8
Qualitative and Quantitative Detection Limit (IUPAC Method)

| Sample No. | Rbl | 0.05 µg/mL | 0.10 µg/mL |
|------------|-------|------------|------------|
| | PA | PA | PA |
| 1 | 1.07 | 6.32 | 13.66 |
| 2 | 1.14 | 5.91 | 13.04 |
| 3 | 0.87 | 6.39 | 13.19 |
| 4 | 1.24 | 5.98 | 13.32 |
| 5 | 1.25 | 5.81 | 13.35 |
| 6 | 1.21 | 6.38 | 13.96 |
| N | 6 | 6 | 6 |
| Mean | 1.13 | 6.13 | 13.42 |
| Std Dev | 0.14 | 0.26 | 0.34 |
| CV | 0.128 | 0.042 | 0.025 |

PA = Integrated Peak Area (SO₄²⁻)/1,000,000
Rbl = Reagent Blank

Using the equation: $C_{ld} = k(sd)/m$

Where:

C_{ld} = the smallest reliable detectable concentration an analytical instrument can determine at a given confidence level.

k = 3 (Qualitative Detection Limit, 99.86% Confidence)

= 10 (Quantitative Detection Limit, 99.99% Confidence)

sd = standard deviation of the reagent blank (Rbl) readings.

m = analytical sensitivity or slope as calculated by linear regression.

$C_{ld} = 3(0.14)/22.45 = 0.0187$ µg/mL as SO₄²⁻ for the qualitative limit.

$C_{ld} = 10(0.14)/22.45 = 0.0624$ µg/mL as SO₄²⁻ for the quantitative limit.

Qualitative detection limit = 0.187 µg SO₄²⁻ (10-mL sample volume) or 0.005 ppm SO₂ (12-L air volume).

Quantitative detection limit = 0.624 µg SO₄²⁻ (10-mL sample volume) or 0.016 ppm SO₂ (12-L air volume).

Table 9
Summary - Comparison of Methods for SO₂

| Set # | Method | SO2 Conc (ppm) | N | Std Dev |
|-------|--------|----------------|---|---------|
| 1 | THE | 1.36 | | |
| | 104M | 1.36 | 6 | 0.156 |
| | IABC | 1.35 | 4 | 0.057 |
| 2 | THE | 2.40 | | |
| | 104M | 2.31 | 6 | 0.058 |
| | IABC | 2.34 | 6 | 0.088 |
| 3 | THE | 4.40 | | |
| | 104M | 4.16 | 5 | 0.358 |
| | IABC | 3.93 | 6 | 0.214 |
| 4 | THE | 5.32 | | |
| | 104M | 5.79 | 6 | 0.497 |
| | IABC | 5.35 | 4 | 0.152 |

Notes: (a) THE = Theoretical (Taken) value, calculated from certified SO₂ cylinder and gas generation system flows.

(b) 104M = Impinger samples taken using OSHA Method No. ID-10 (modified).

(c) IABC = Impregnated activated beaded carbon

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Table 10
 Sampling and Analysis - Type II Sampling Tube
 (1×TWA-PEL, 25 °C and 80% RH)

| Sample No. | Air Vol (L) | Found | ppm SO ₂ | | Statistical Analysis | | | Recovery (%) |
|------------|-------------|-------|---------------------|---|----------------------|---------|-------|--------------|
| | | | Taken | N | Mean | Std Dev | CV | |
| 1 | 10.6 | 1.65* | 2.23 | 7 | 2.09 | 0.11 | 0.052 | 93.6 |
| 2 | 10.3 | 2.08 | 2.23 | | | | | |
| 3 | 11.3 | 2.23 | 2.23 | | | | | |
| 4 | 12.7 | 2.16 | 2.23 | | | | | |
| 5 | 10.6 | 1.91 | 2.23 | | | | | |
| 6 | 10.3 | 2.14 | 2.23 | | | | | |
| 7 | 11.3 | 1.98 | 2.23 | | | | | |
| 8 | 12.7 | 2.10 | 2.23 | | | | | |

* Outlier, not used in statistical analysis

Table 11
 Comparison Study - With/Without Pre-filter
 (1×TWA-PEL, 25 °C and 50% RH)

| Sample Set # | With Pre-filter | | Without Pre-filter | |
|--------------|-----------------|---------------------|--------------------|---------------------|
| | Air Vol (L) | ppm SO ₂ | Air Vol (L) | ppm SO ₂ |
| 1 | 12.1 | 1.80 | 11.8 | 1.79 |
| 2 | 12.1 | 1.62 | 11.1 | 1.71 |
| 3 | 11.2 | 2.04 | 12.3 | 1.96 |
| 4 | 11.8 | 1.92 | * | |
| 5 | 11.1 | 1.79 | 12.1 | 1.84 |
| 6 | 12.3 | 1.89 | 11.2 | 1.95 |
| N | | 6 | | 5 |
| Mean | | 1.84 | | 1.85 |
| Std Dev | | 0.14 | | 0.11 |
| CV | | 0.077 | | 0.058 |

* Pump stopped during sampling
 Notes: (a) Sampling Time = 120 min
 (b) Flow Rate ≈ 0.1 L/min
 (c) Sample Solution Volume for Desorption = 10 mL

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Block Diagram of the Laboratory Generation System

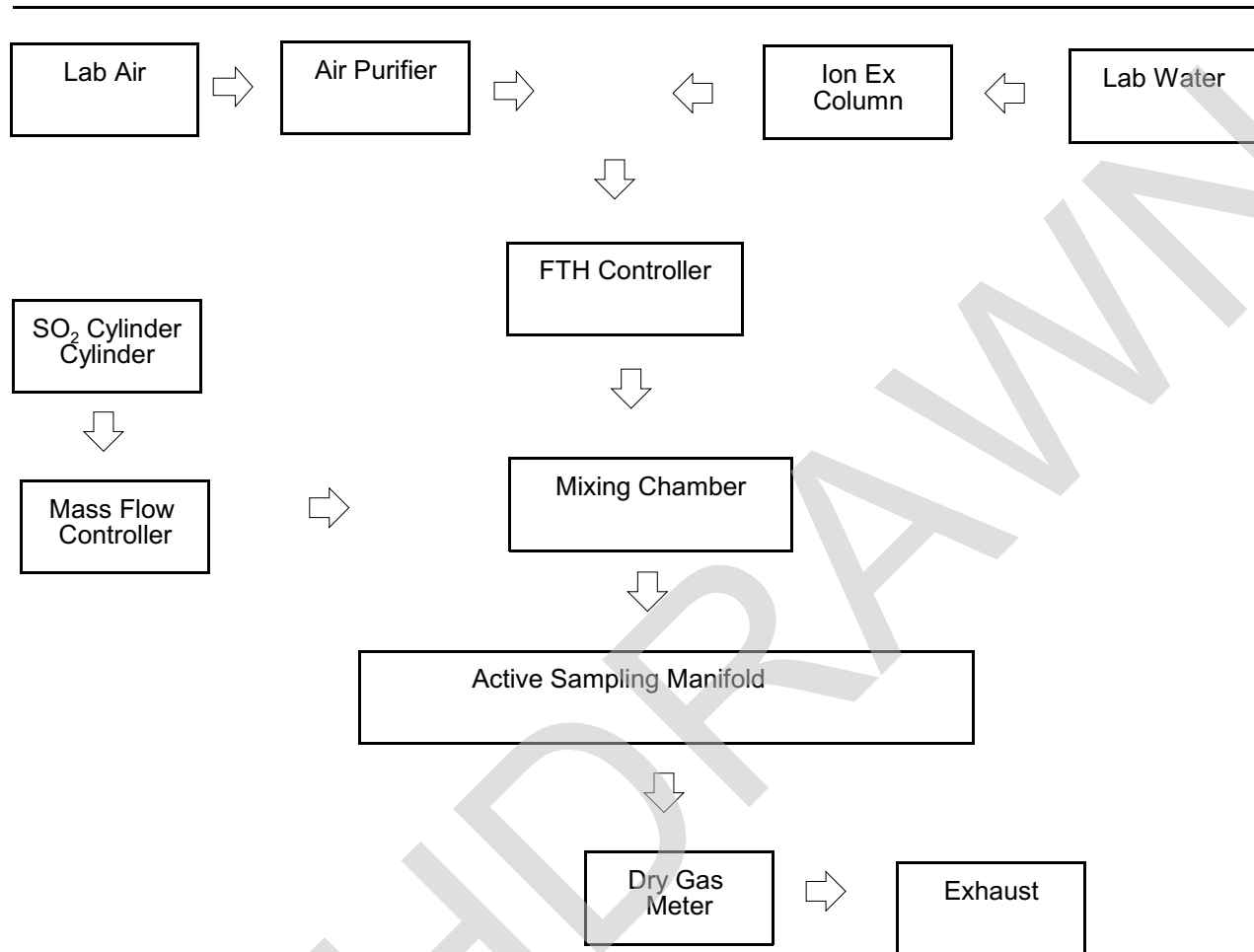


Figure 1

The system shown above provided a means for generating dynamic test atmospheres. The system consists of four essential elements:

- 1) a flow-temperature-humidity (FTH) control system,
- 2) an SO₂ vapor generating system,
- 3) a mixing chamber, and
- 4) an active sampling manifold.

Note: OSHA no longer uses or supports this method (December 2019).